

# Recap of Consultation Meetings and some simulations

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18<sup>th</sup> September 2025  
LhARA Collaboration Meeting 8

# Consultation Meeting

- As part of PA1 WP5 held 3 user consultation meetings
  - Online (Ruth hosted, stage 1)
  - Birmingham (Tony hosted, stage 1)
  - Liverpool (Narender/Milaan hosted, stage 2)
- Aim to draw a list of requirements for the facility in order to allow a wide range of users to optimise their work
- Main focus Stage 1 in-vivo but also Stage 2 in-vitro and companion animals

# Consultation Meetings Conclusions

## Conclusions

- C1:** The case for a change to the present baseline beam-delivery concept for the low and high energy *in-vitro* end stations and the *in-vivo* end station is not compelling and therefore the present baseline should be retained.
- C2:** A specification of 5% as the upper limit on the accuracy of the integrated dose measurement and its repeatability is sufficient for the dose-measurement uncertainty not to dominate the error budget of biological experiments.
- C3:** Any setup and end-station must be **“Simple, Robust, Reproducible, and Cheap”**.
- C4:** For the rest of the consultation process, the Stage 1 *in-vitro* experiments will assume the use of standard plastic cell dishes.
- C5:** An X-ray source to be included in the facility to allow control sample and low LET comparisons to be made with cultures in both the stage 1 and stage 2 *in-vitro* end-stations
- C6:** Integration of cell transport into the end-stations, and environmental stabilisation needs to be in the order of minutes to ensure cell viability.
- C7:** Temperature and oxygen stability must be maintained and monitored and be set for user requirements.
- C8:** The experimental complications arising from using a low-energy proton beam must be considered carefully and a Geant4 simulation has been developed to extract kinetic Energy, LET, profiles etc.

# Consultation Meetings Conclusions

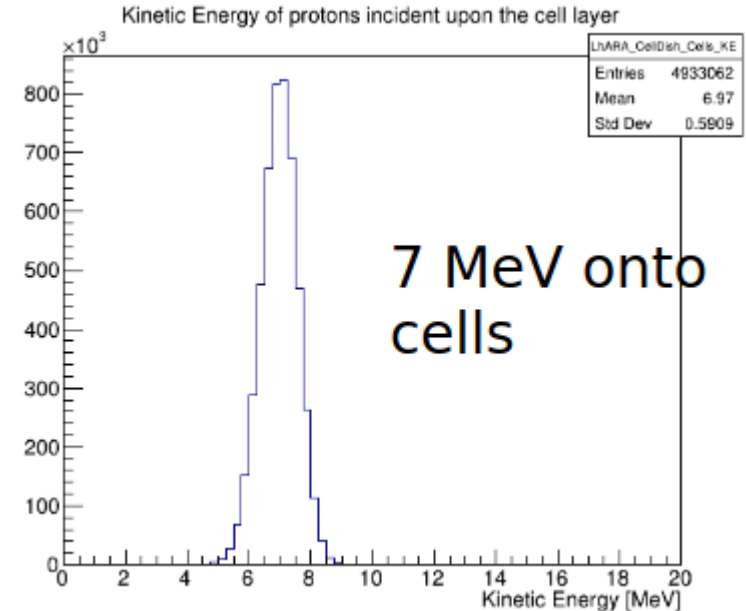
## Conclusions

- C9:** Animal house on site is essential for animal work as it is a "One-way trip" for animals involved in experiments.
- C10:** Support will be required from external experts regarding the regulations for animal work. Development of the specification of the *in-vitro* end station and its operation should include careful consideration of the range of animals required. Mary Lyons have been involved thus far.
- C11:** Real time imaging will be required in the end-station room for animal work, alignment, reproducibility e.g cone-beam CT.

# Pre-CDR Geometry

**Simulation setup to closely match that of the Pre-CDR.**

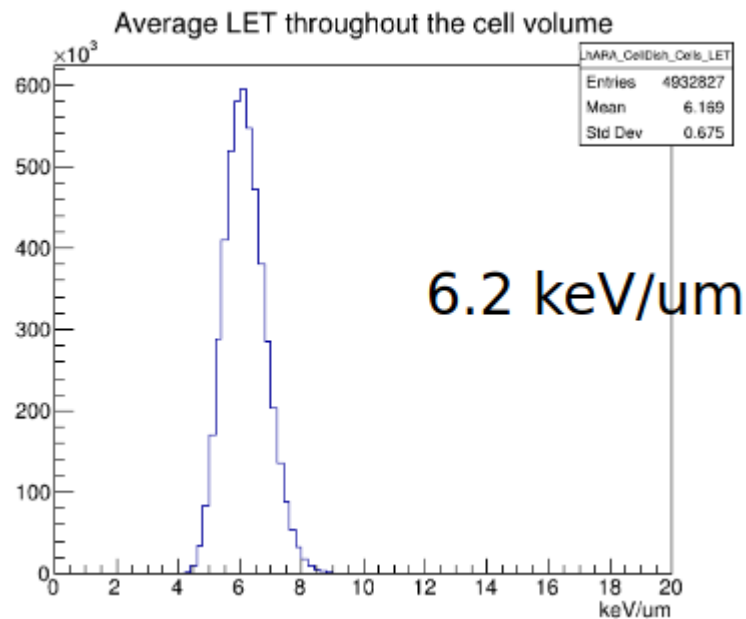
- \* **25um Ti vacuum window**
- \* **100um end-station plastic window**
- \* **250um plastic beam monitor**
- \* **5mm air gap**
- \* **1.3 mm cell dish base**
- \* **30um cells**
- \* **2mm water (or Marcus chamber for dose)**



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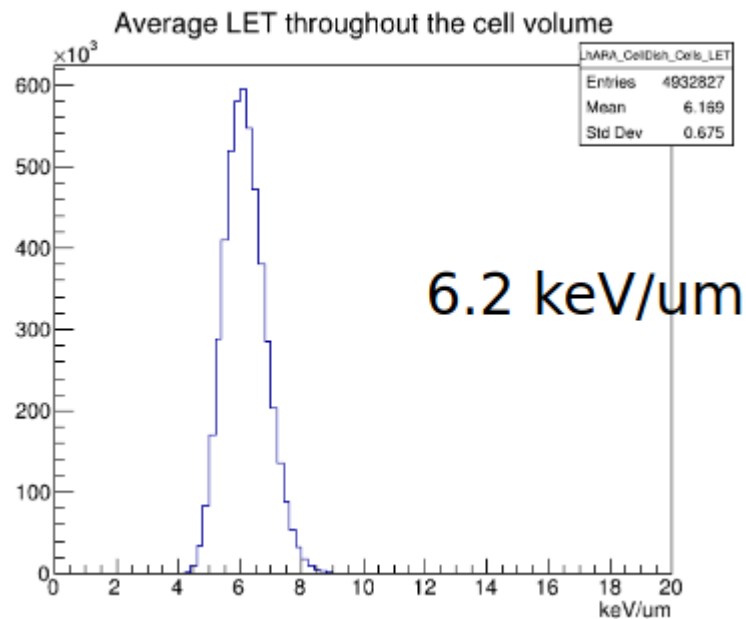
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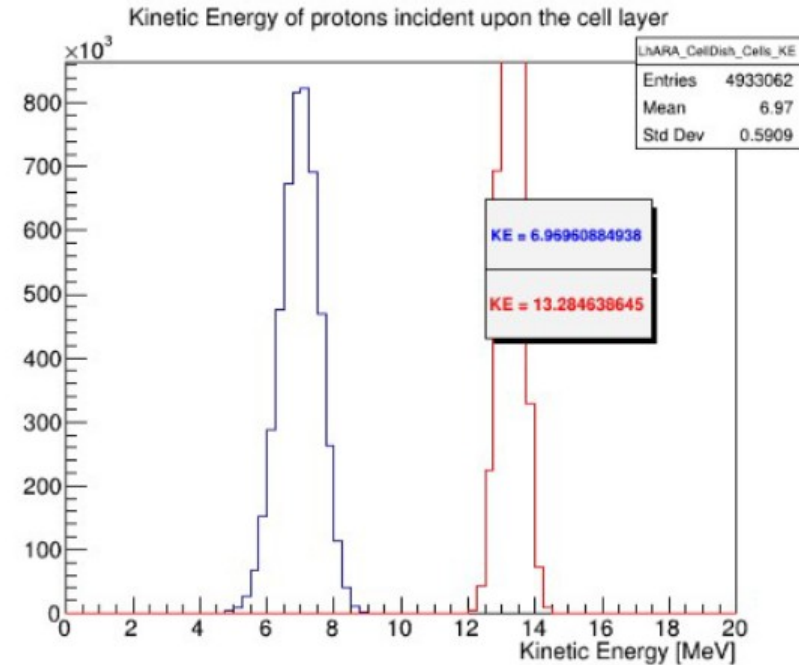
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- \* **25um Ti vacuum window**
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- \* **2.5 um Mylar dish base**
- \* **30um cells**
- \* **2mm water (or Marcus chamber for dose)**

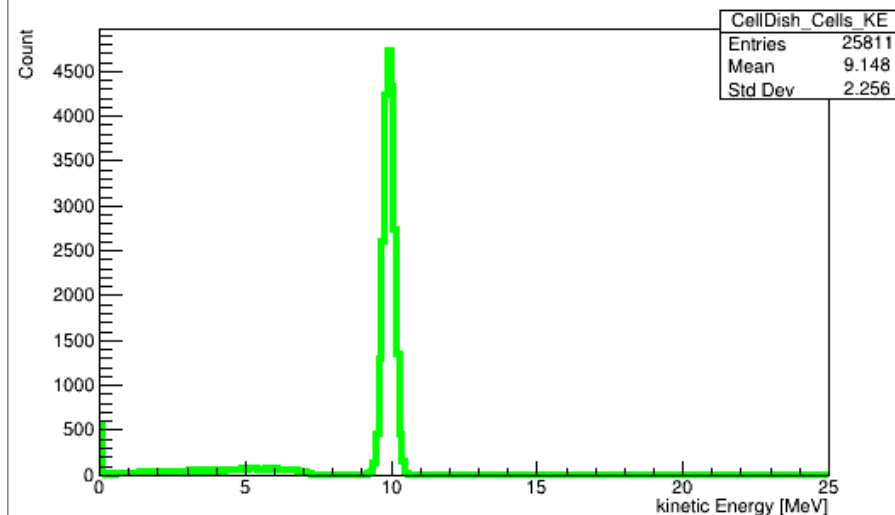
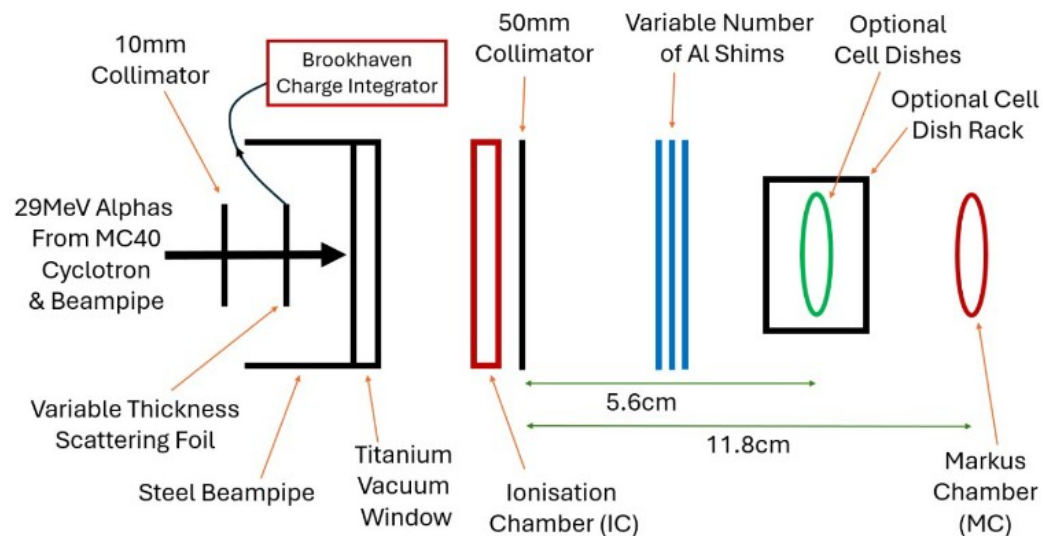
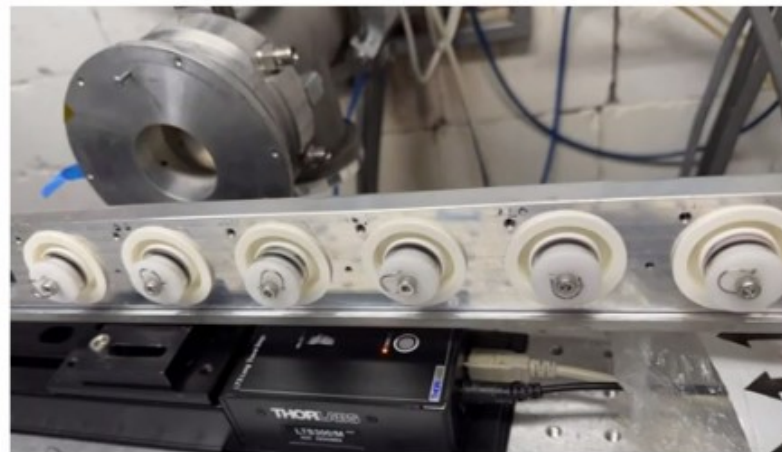




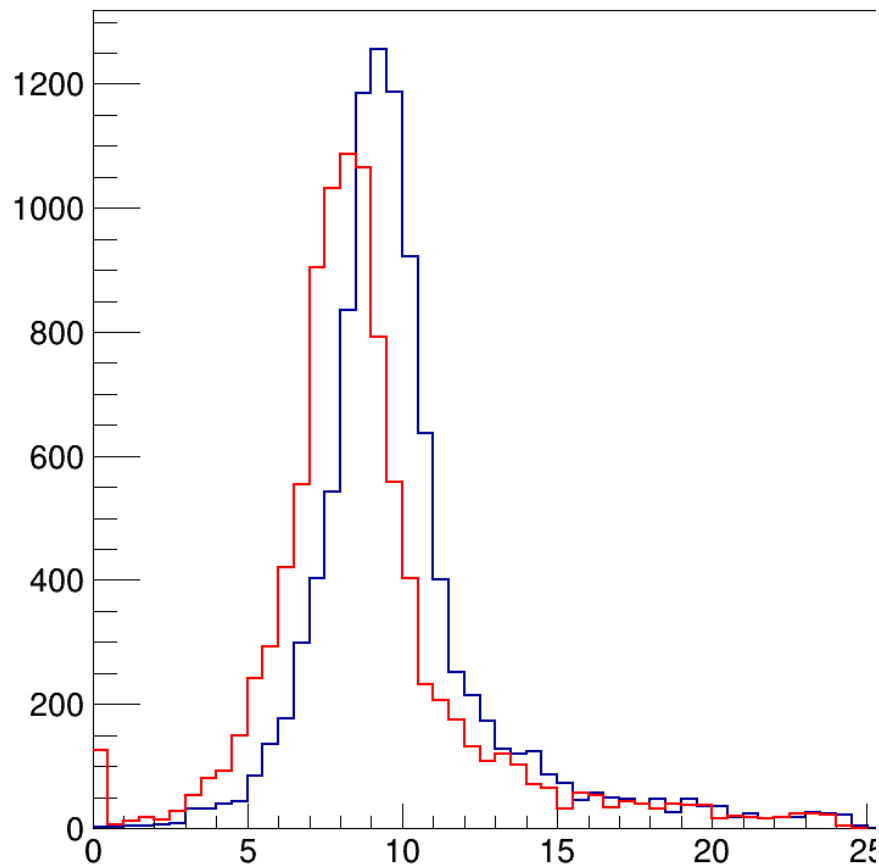
## Radiobiology Preparations 2 – Protons (14 vs 28 MeV)

## Pre-plating protocol:

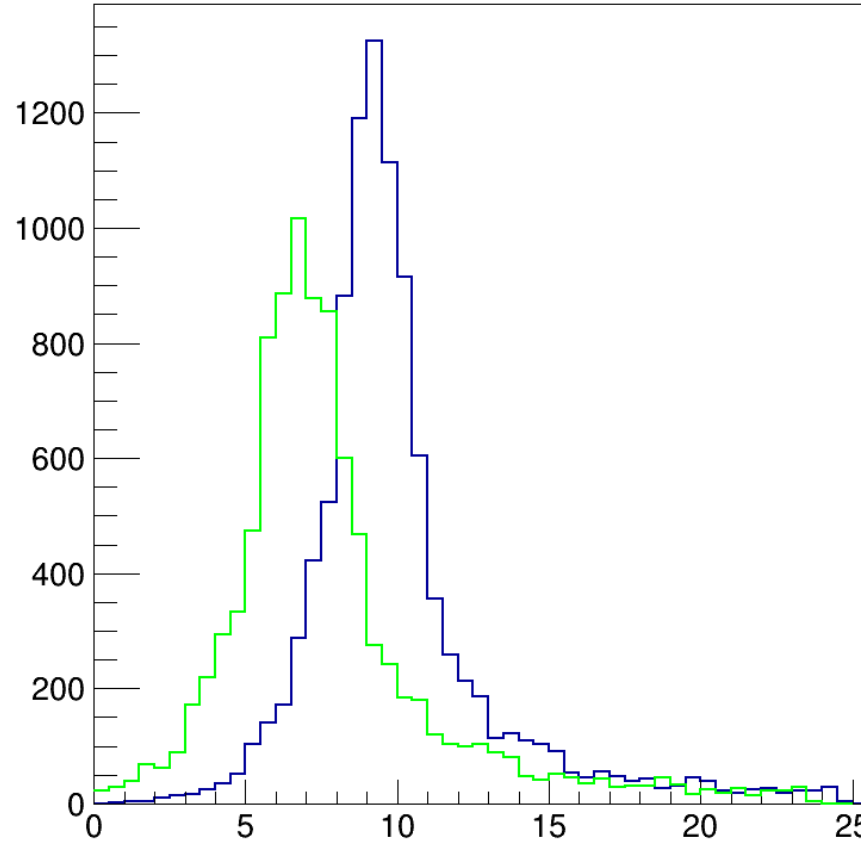
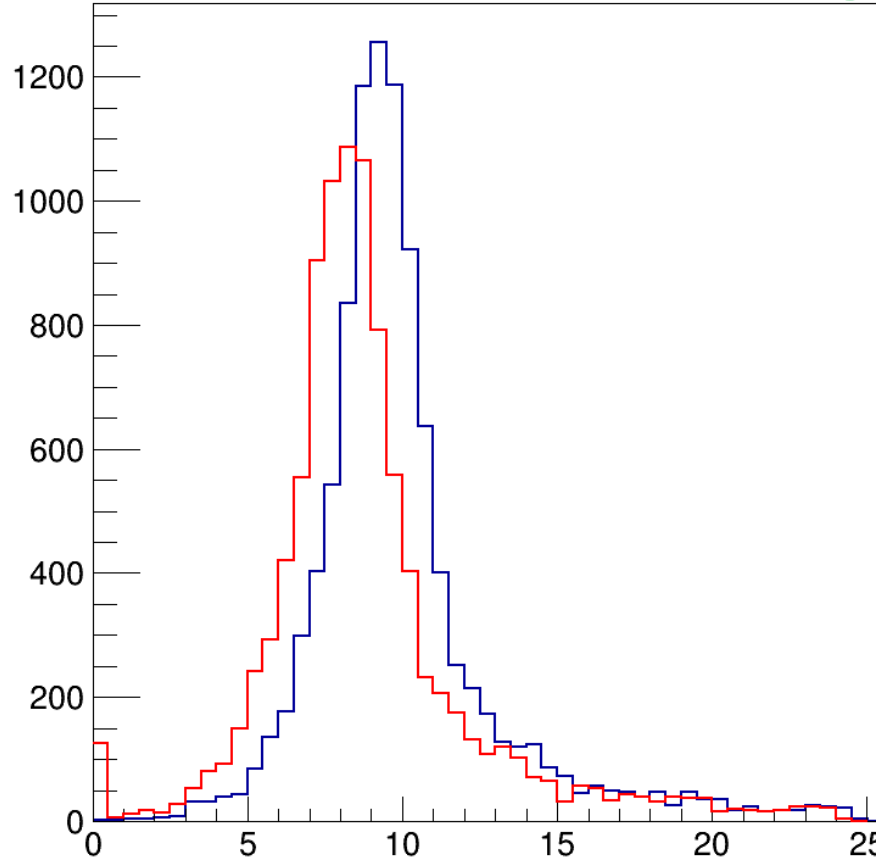
1. Seed single cells on mylar the day before (PM)
2. **Remove media** and Irradiate single cells **vertically**
3. Change media
4. Grow colonies for 7-14 days
5. Stain and count



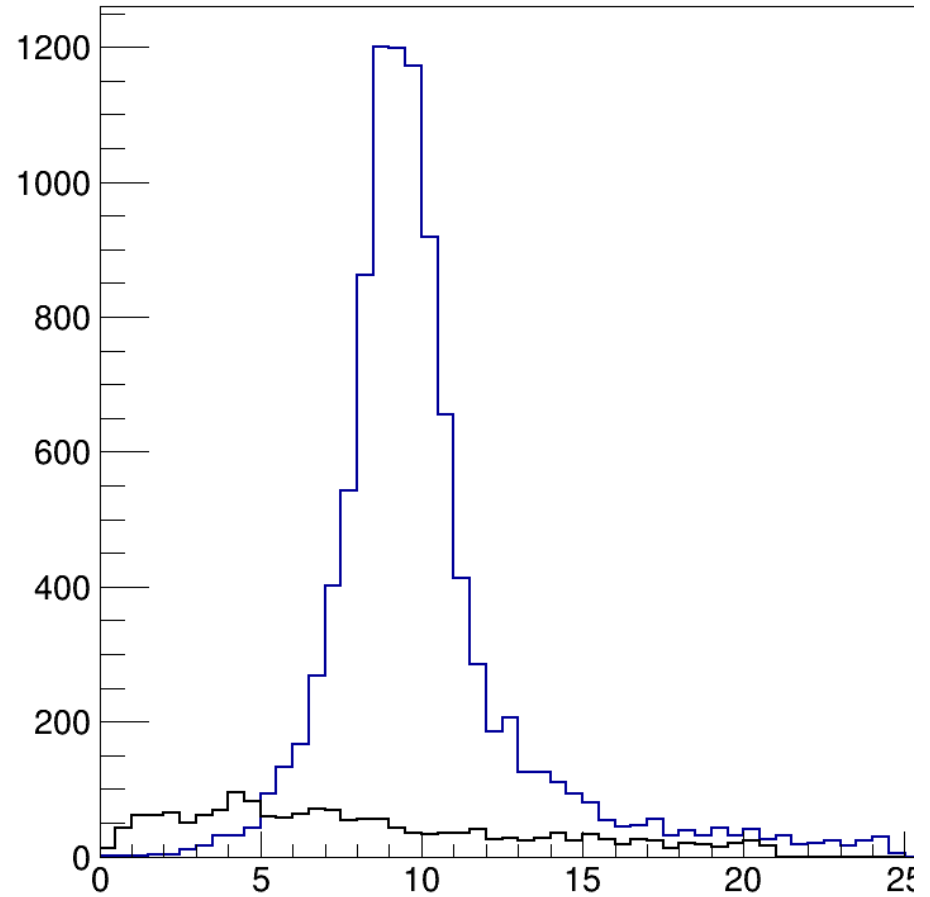
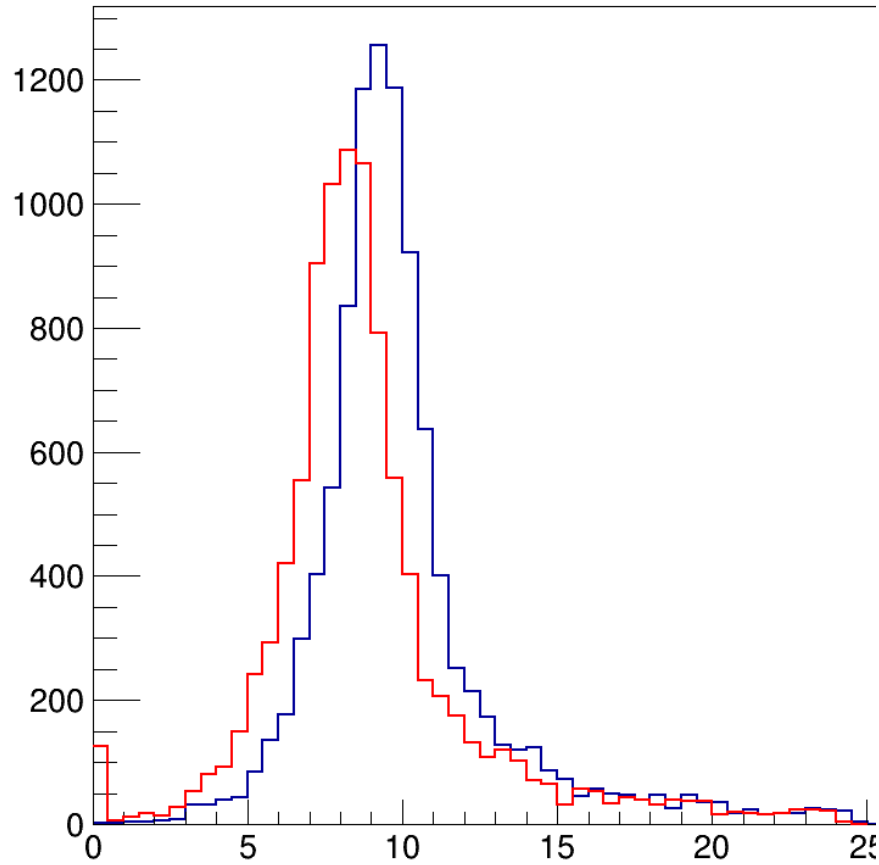
# SCAPA, 2.5um Mylar



# SCAPA, 2.5um Mylar (+) 130um Glass



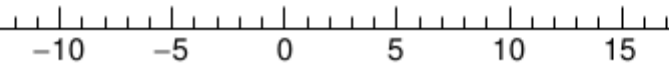
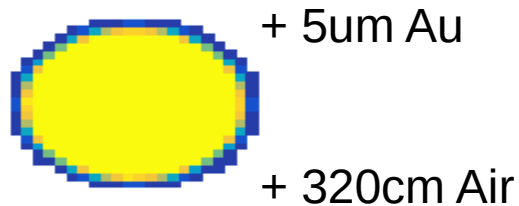
# SCAPA, 1.2mm Polystyrene



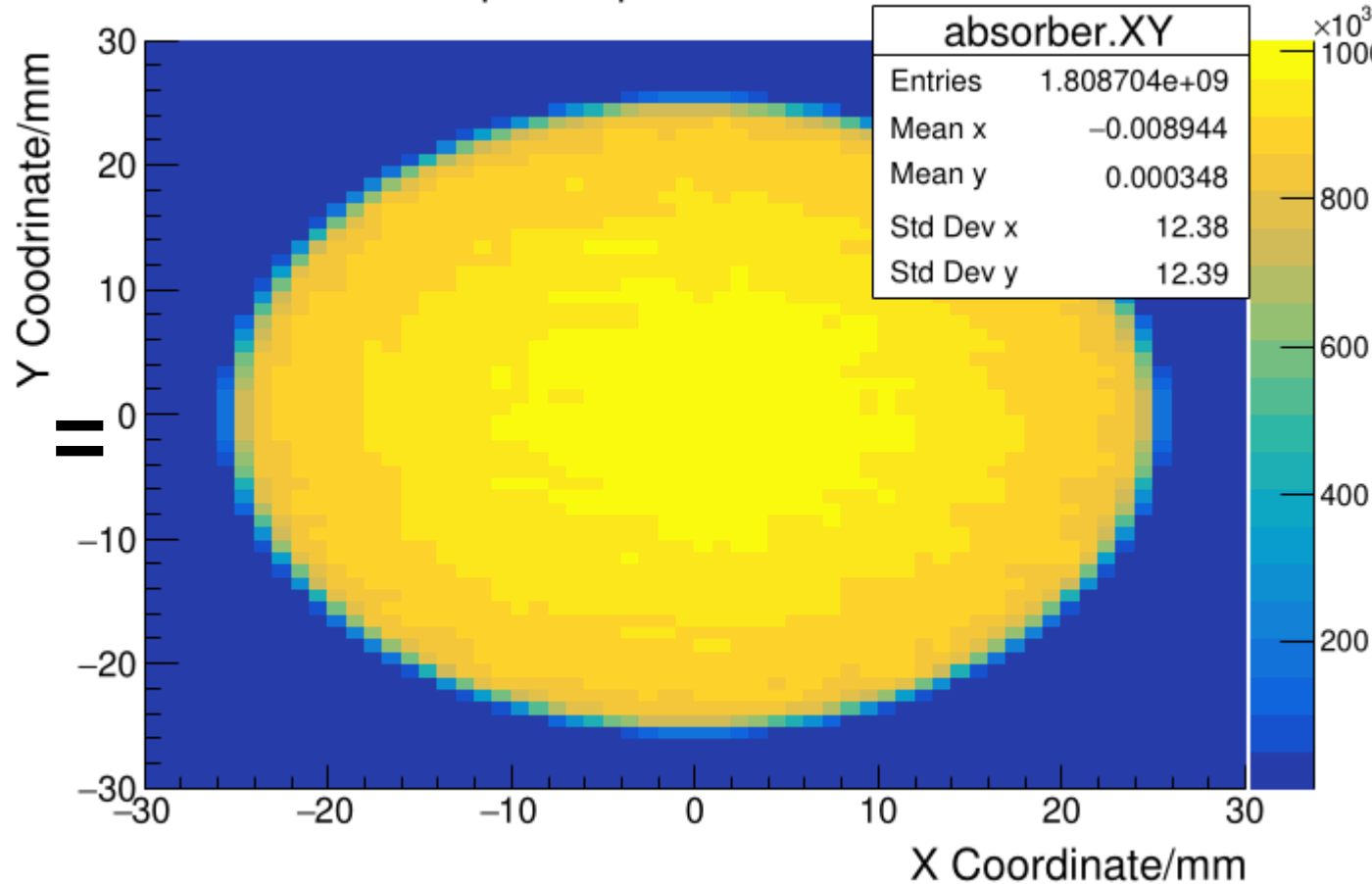
# Scattering Foil – 28 MeV alpha

## Geant4

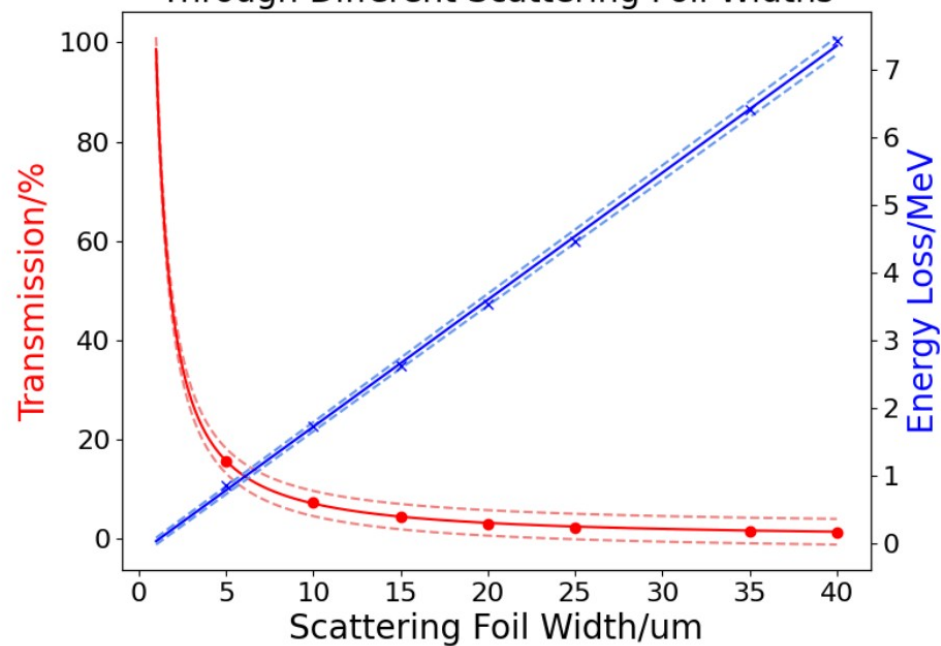
plot of generated particle position



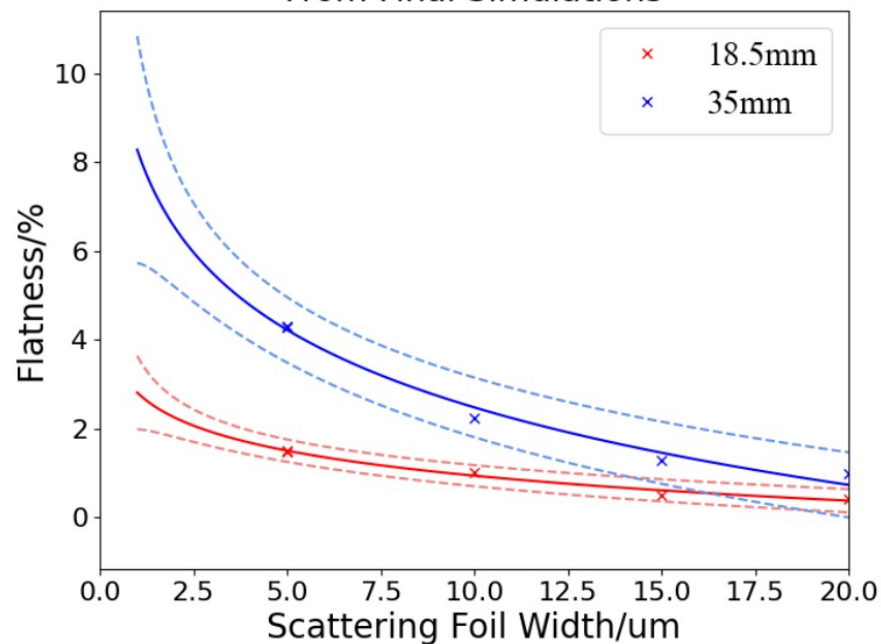
Scatter plot of particles into absorber



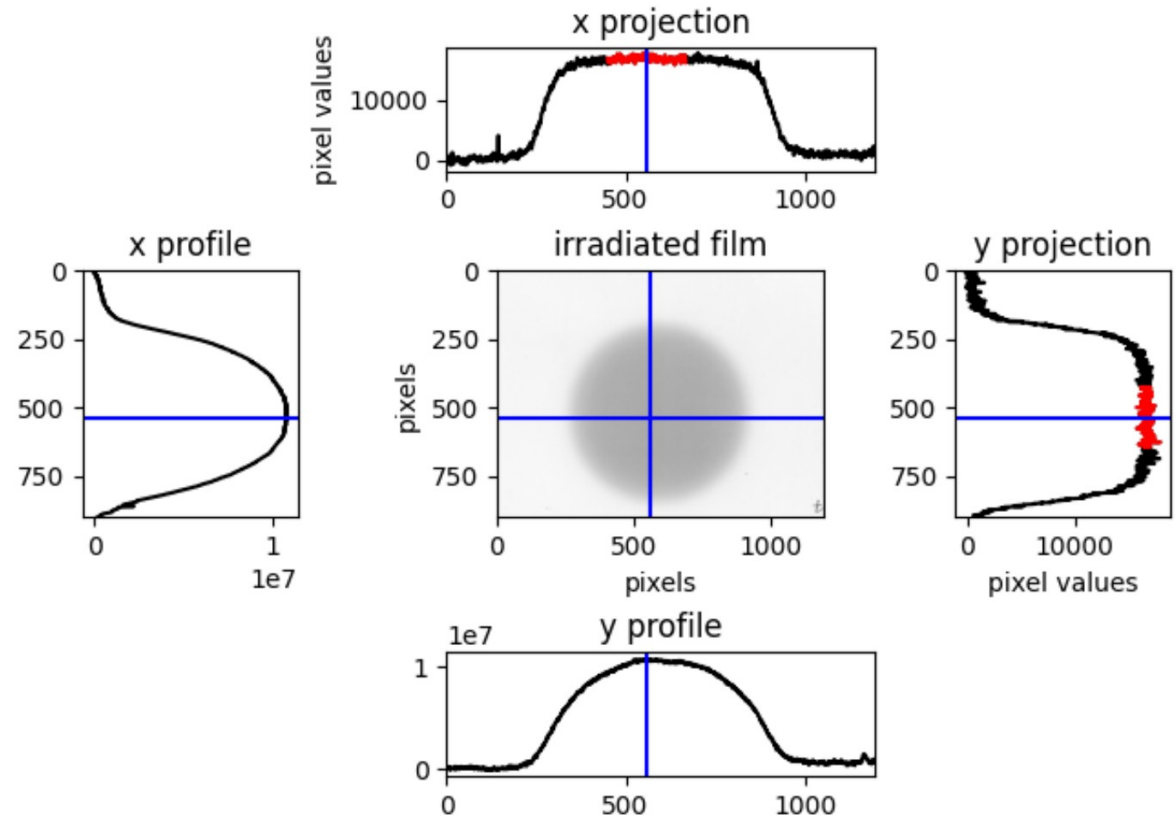
Energy loss and Transmission of a 29MeV Alpha Beam Through Different Scattering Foil Widths



Plot of Flatness against Scattering Foil Width From Final Simulations



# Scattering Foil – 28 MeV alpha data



# Scattering Foil – 28 MeV alpha data

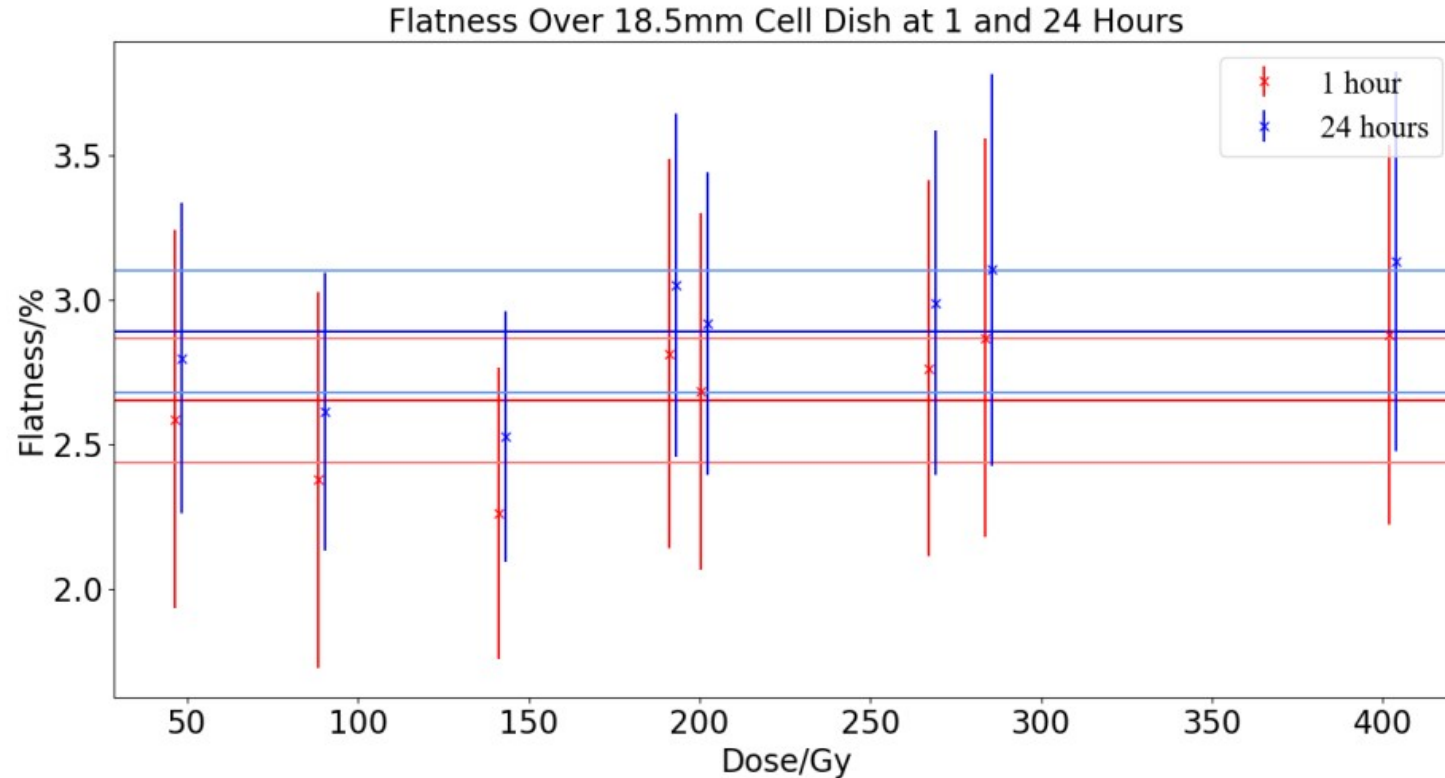


Figure 4.6: Flatness measured over 18.5mm at 1 and 24 hours for each film